

Calculation of optimum number of enzyme units

$$\frac{\text{length of your DNA [bp]}}{\text{number of restriction sites}} = \text{your site frequency [bp]}$$

$$\frac{48502 \text{ bp}}{\text{number of } \lambda \text{ restriction sites}} = \lambda \text{ site frequency [bp]}$$

$$\frac{\lambda \text{ site frequency}}{\text{your site frequency}} = \text{number of units for 1 } \mu\text{g DNA} = X_1$$

$$X_1 \times \text{mass of DNA } [\mu\text{g}] = \text{number of units needed for digestion}$$

If the enzyme doesn't cut λ DNA, often Adenovirus 2 DNA is used, which is 35937 bp long.

Example:

5 μg of a 5000 bp plasmid carrying 5 enzyme sites shall be digested.

→ your site frequency = $5000 / 5 = 1000$

The same enzyme cuts phage λ DNA also 5 times.

→ λ site frequency = $48502 / 5 = 9700 \approx 10000$

→ number of units for digestion of 1 μg DNA = $10000 / 1000 = 10$

→ number of units for digestion of 5 μg DNA = $10 \times 5 = 50$

- Supercoiled plasmids need more unit activity (up to 5x more) for complete cleavage compared to linearized DNA.
- Use a maximum of 10% enzyme in the total reaction volume, otherwise the glycerol will inhibit the reaction.
- All of Roche's enzymes have a concentration of 10 units/ μl

Length of your DNA [bp]	Sites in your DNA	Sites in λ DNA
	1	
Amount of DNA [μg]	Units needed	
1	#DIV/0!	

Length of your DNA [bp]	Sites in your DNA	Sites in Ad2 DNA
	1	
Amount of DNA [μg]	Units needed	
1	#DIV/0!	

<https://www.roche-applied-science.com/sis/cloning/cloning.jsp?id=010103>

Examples of enzymes assayed with λ DNA:

ApaI: 1 site (prev. dig. with HindIII (6 sites))
 EcoRV: 21 sites
 HpaI: 14 sites
 KpnI: 2 sites
 MunI: 8 sites
 NdeI: 7 sites
 NheI: 1 site (prev. dig. with EcoRI (5 sites))
 ScaI: 5 sites
 StuI: 6 sites
 XhoI: 1 site

Examples of enzymes assayed with Ad2 DNA:

NotI: 7 sites
 SpeI: 3 sites